

## Self-Assembly

**Kinetic Self-Assembly: Selective Cross-Catenation of Two Sterically Differentiated Pd<sup>II</sup>-Coordination Rings\*\****Akiko Hori, Ken-ichi Yamashita, and Makoto Fujita\**

Like protein folding, self-assembly in biology is considered to be a kinetic process where the most favorable pathway to programmed structures is kinetically selected through meta-stable intermediary structures.<sup>[1–3]</sup> Although numerous reports have appeared on thermodynamically controlled self-assembly,<sup>[4–8]</sup> relatively few have discussed kinetic self-assembly in chemical systems.<sup>[9–13]</sup> Here we deal with kinetic self-assembly based on the dynamic behavior of coordination rings. We emphasize that designing not only structures but also pathways is an important yet unexplored task for controlling molecular self-assembly.

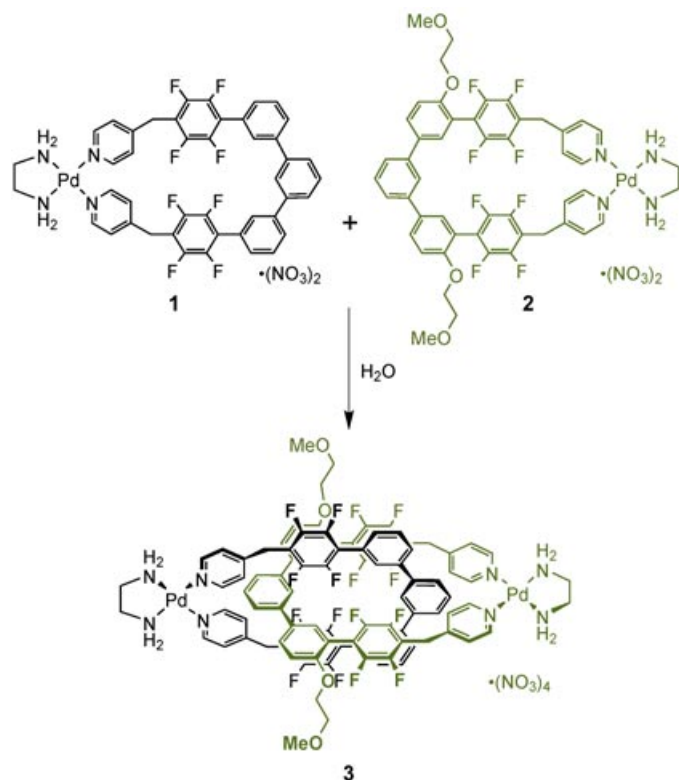
The kinetic self-assembly discussed here is demonstrated by the cross-catenation<sup>[14]</sup> of Pd<sup>II</sup>-linked rings **1** and **2**, which

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[\*\*] This research was supported by the CREST project of the Japan Science and Technology Corporation (JST), for which M.F. is the principal investigator, and also, in part, by Genesis Research Institute, Inc., to which A.H. is thankful for a fellowship.

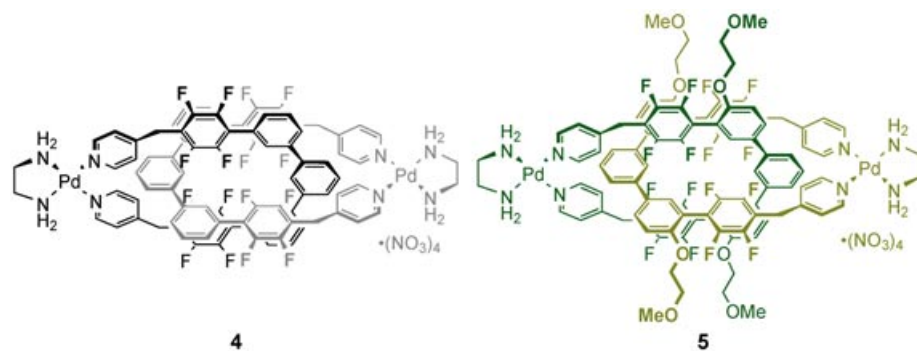
are differentiated by alkoxy side chains (Scheme 1). The homocatenation of **2** is kinetically unfavorable because of the steric demand of the alkoxy side chains.<sup>[15]</sup> Accordingly, ring **2**



Scheme 1.

is only allowed to thread on less-hindered **1**, thus giving rise to the kinetic formation of cross-catenane **3**.

We first estimated the timescale of the homocatenation of **1** by 2D EXSY NMR spectroscopy.<sup>[16]</sup> This sterically less hindered ring rapidly forms catenanes,<sup>[17]</sup> and a clear correlation between **1** and its catenated dimer **4** was observed even after 30 ms mixing time, thus showing that the catenation takes place on a millisecond timescale. The rapid catenation should be retarded if a sterically demanding side chain is attached to the ligand of **1**. A bis(methoxyethoxy)-substituted ligand (the precursor of **2**) was synthesized and complexed



with  $[\text{Pd}(\text{en})(\text{NO}_3)_2]$  (en = ethylenediamine) in DMSO to give **2** quantitatively. Since the catenation of  $\text{Pd}^{\text{II}}$ -linked rings is driven by aromatic-stacking interactions, which work efficiently in aqueous media,<sup>[17]</sup> water was added to the solution of **2** in DMSO to transform it into homocatenane **5**. As expected, the catenation was significantly retarded, and took 4–5 h to be completed (Figure 1).

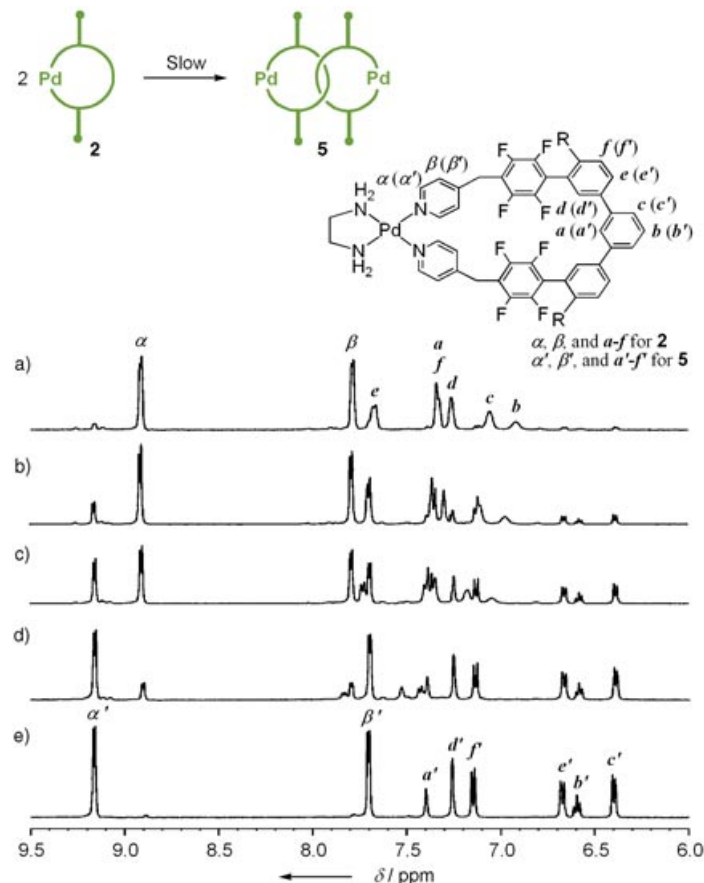
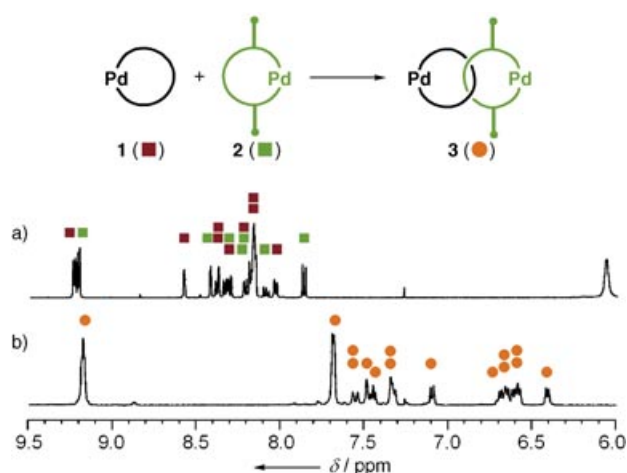


Figure 1. NMR spectra indicating the transformation of **2** into catenane **5** (aromatic region, 500 MHz, 300 K,  $[\text{D}_6]\text{DMSO}/\text{D}_2\text{O}$  (1:1.5), TMS as an external standard); after a) 3 min, b) 10 min, c) 20 min, d) 60 min, e) 240 min.

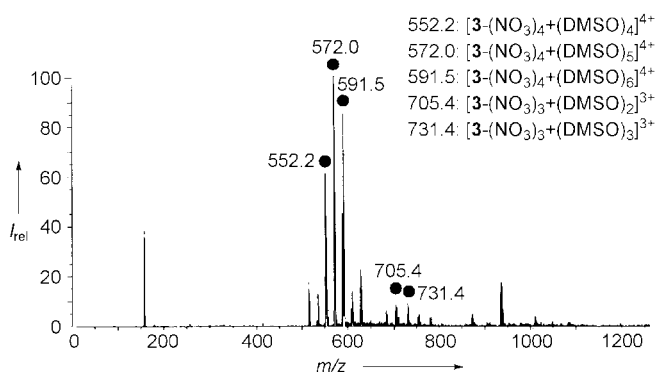
Accordingly, we examined the cross-catenation of **1** and **2** by utilizing the large difference in the catenation rate of **1** and

**2**. Water (0.9 mL) was added to a solution of **1** (4.5 mg, 4.5  $\mu\text{mol}$ ) and **2** (5.2 mg, 4.5  $\mu\text{mol}$ ) in DMSO (0.6 mL) at ambient temperature. The two rings were immediately transformed into a single catenated product as indicated by NMR spectroscopic analysis (Figure 2). The large upfield shifts of the central aromatic protons are characteristic of interstrand stacking between the rings.<sup>[18]</sup> Since this spectrum was different from those of homocatenated compounds **4** and **5**, the selec-



**Figure 2.** Observation of cross-catenation by NMR spectroscopy (aromatic region, 500 MHz, 300 K, TMS as an external standard): a) The spectrum of a mixture of **1** and **2** in  $[D_6]DMSO$ , b) the spectrum after the addition of  $D_2O$ .

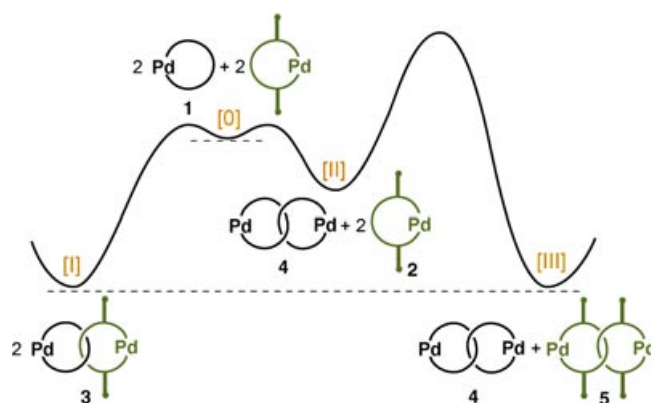
tive formation of cross-catenane **3** was strongly suggested. A coldspray ionization mass spectrometry (CSI-MS)<sup>[19]</sup> study confirmed the selective formation of cross-catenane **3** as evident from prominent signals that fully agreed with **3**: for example,  $m/z$  572.0  $[3-(NO_3)_4+(DMSO)_5]^{4+}$  (Figure 3). Signals indicating the presence of **4** or **5** were barely visible.



**Figure 3.** CSI mass spectrum of cross-catenane **3** in  $DMSO/H_2O$  (1:1.5) solution.

The selective formation of **3** is explained by the energy diagram shown in Figure 4. Although three possible catenanes **3–5** have similar thermodynamic stability, the formation of **5** is kinetically inhibited because ring **2** must thread on the bulky ligand of **2** itself. Thus, the states [I] and [II] will rapidly equilibrate and favor the formation of cross-catenane **3**.

This speculation suggests that cross-catenane **3** (state [I]) will be equilibrated with **4** + **5** (state [III]) if the reaction time is long (or temperature is high) enough to promote the formation of **5**. In fact, when the solution of **3** was allowed to stand at 25 °C for a week, new signals assigned to homocatenanes appeared in the NMR spectrum. The ratio of **3**:**4**:**5** was estimated to be 4.3:1:1 and 2.8:1:1 after 1 day and 2 days, respectively. The ratio finally became statistical (2:1:1) after



**Figure 4.** The energy diagram for the homo- and cross-catenation of **1** and **2**.

8 days. The same statistical mixture was also obtained when presynthesized catenanes **4** and **5** were combined in a mixture of  $H_2O$  and  $DMSO$ , and the solution was allowed to stand at 25 °C for 1 week. Clearly, the selective formation of cross-catenane over short times (minutes or hours) is kinetically controlled while the formation of a statistical mixture over longer times (days) is thermodynamically controlled. Of course, the equilibration became much slower at lower temperatures. The transformation of **3** into a statistical mixture took several months in a refrigerator (6 °C). Moreover, this transformation was completely suppressed in a freezer.

In summary, we have controlled the cross-catenation of two different rings by a kinetic method. Although still a preliminary study, the case detailed here demonstrates a new method of molecular self-assembly in which desired structures are generated by *programmed pathways*. Our results offer a good example of such kinetically controlled self-assembly and will, hopefully, be emulated by many other cases.

Received: May 15, 2004

**Keywords:** catenanes · molecular recognition · palladium · self-assembly

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- [18] Selected data for **3**: <sup>1</sup>H NMR (500 MHz, [D<sub>6</sub>]DMSO/D<sub>2</sub>O = 1:1.5, 280 K, TMS as an external standard): δ = 9.23 (s, 8H, H<sub>α</sub> and H<sub>α'</sub>), 7.74 (s, 8H, H<sub>β</sub> and H<sub>β'</sub>), 7.61 (s, 1H, H<sub>a</sub>), 7.59 (s, 1H, H<sub>a'</sub>), 7.54 (s, 2H, H<sub>d</sub>), 7.45 (t, *J* = 6.2 Hz, 2H, H<sub>f</sub>), 7.39 (s, 2H, H<sub>f'</sub>), 7.33 (d, *J* = 6.2 Hz, 2H, H<sub>g</sub>), 7.11 (d, *J* = 8.0 Hz, 2H, H<sub>f</sub>), 6.72 (t, *J* = 7.2 Hz, 1H, H<sub>g'</sub>), 6.69 (d, *J* = 6.2 Hz, 2H, H<sub>c</sub>), 6.64–6.58 (m, 5H, H<sub>b</sub>, H<sub>c</sub>, H<sub>c'</sub>), 6.45 (d, *J* = 7.2 Hz, 2H, H<sub>c'</sub>), 4.25 (s, 4H, CH<sub>2</sub>), 4.17 (s, 8H, CH<sub>2</sub>, CH<sub>2'</sub>), 3.72 (s, 4H, CH<sub>2</sub>), 3.32 (s, 6H, CH<sub>3</sub>), 3.05 ppm (s, 8H, en, en'); <sup>13</sup>C NMR (125 MHz, [D<sub>6</sub>]DMSO/D<sub>2</sub>O = 1:1.5, 280 K): δ = 155.4 (Cq), 152.1 (Cq), 152.0 (CH<sub>α</sub>, CH<sub>α'</sub>, Cq), 145.5–142.0 (4 × CF), 139.2 (Cq), 138.9 (Cq), 138.4 (Cq), 132.2 (Cq), 129.7 (CH<sub>2</sub>), 129.6 (CH<sub>2</sub>), 129.3, 129.2, 129.0 (CH<sub>b</sub>, CH<sub>b'</sub>, CH<sub>c'</sub>), 128.1 (CH<sub>β</sub>, CH<sub>β'</sub>), 127.8 (CH<sub>d'</sub>), 127.4 (Cq), 127.0 (CH<sub>c</sub>), 126.4 (CH<sub>d</sub>), 125.1 (CH<sub>c</sub>), 124.2 (CH<sub>c'</sub>), 123.3 (CH<sub>a</sub>), 122.9 (CH<sub>a'</sub>), 118.4 (Cq), 116.4 (Cq), 115.4–115.2 (3 × Cq), 113.9 (CH<sub>f</sub>), 70.4 (CH<sub>h</sub>), 68.5 (CH<sub>g</sub>), 58.4 (CH<sub>i</sub>), 46.9 (en, en'), 27.7 ppm (CH<sub>2</sub> and CH<sub>2'</sub>).
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